Phenology of a relict population of *Wyeomyia smithii* (Diptera:Culicidae) in Tattnall Co., GA

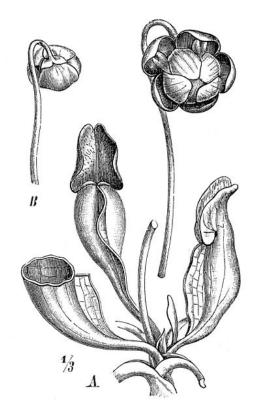
Rachel Morreale, William Irby Georgia Southern University



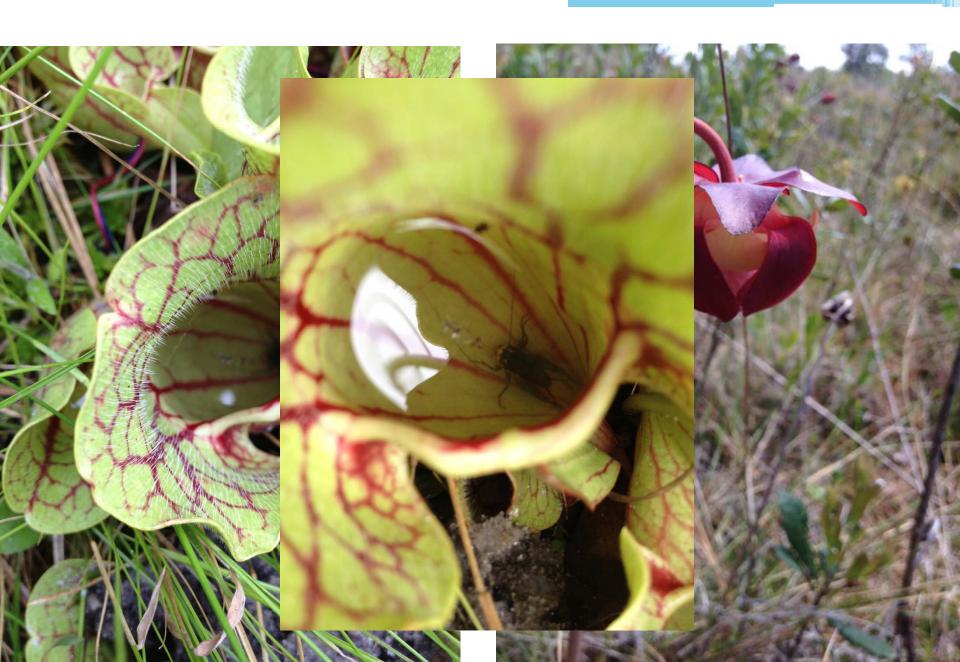
#### Introduction to Sarracenia purpurea

- Range from Gulf of Mexico to Canada
- Grow in rosettes
- Hosts a diverse inquiline community
- Very rare in Georgia



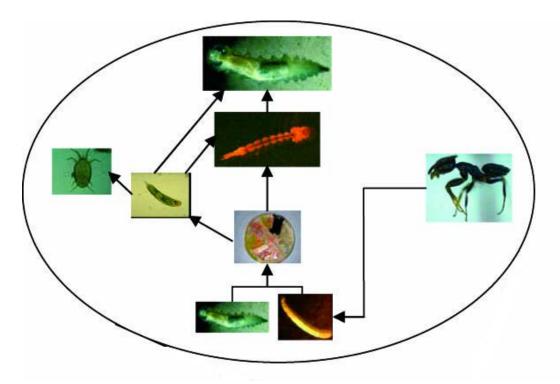


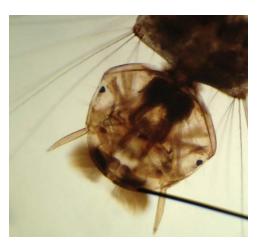




# Introduction to Wyeomyia smithii

- Only found in *Sarracenia purpurea*
- All development occurs inside pitchers
- Nutrient processing chain commensalism





Gotelli and Ellison, 2006.

# Populations differ by latitude

Table 1. Comparison of attributes of Northern and Southern populations of the pitcher-plant mosquito, *Wyeomyia smithii*.

Northern populations

Southern populations

Obligatory autogeny

Univoltine

Winter larval diapause

Facultative autogeny

Multivoltine

Summer and winter larval diapause

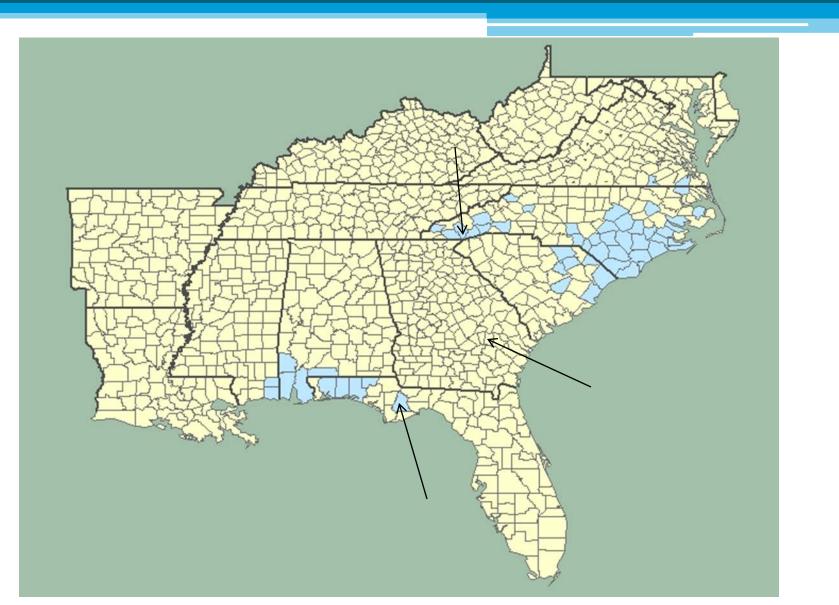


Figure 1. The distribution of S. purpurea in the southeastern United States. Arrows indicate field sites in Florida, Georgia, and North Carolina.

### Site location: Highlands, NC



#### Site Location: Tattnall Co., GA



# Site Location: Apalachicola National



#### Previous Studies: 1997-2000

#### • Behavioral:

 Feeding behavior of post-reproductive females assessed by HIC assay

#### • Reproductive:

Fecundity of autogenous females measured by dissection

#### • Genetic:

- Variation in population genetic structure assessed by isozyme analysis
- Biochemical:
  - Variation in hexamerin expression assessed by SDS-PAGE

**Table 2**: Blood-feeding behavior of adult female Wyeomyiasmithiiafter autogenous egg production during 1998.

Population	Probing	Blood-feeding
Florida	+	+
Georgia	+	* +/-
North Carolina	-	-

\* one female probed, ingested minute quantity of blood; 10 trials, ~50 mosquitoes/trial

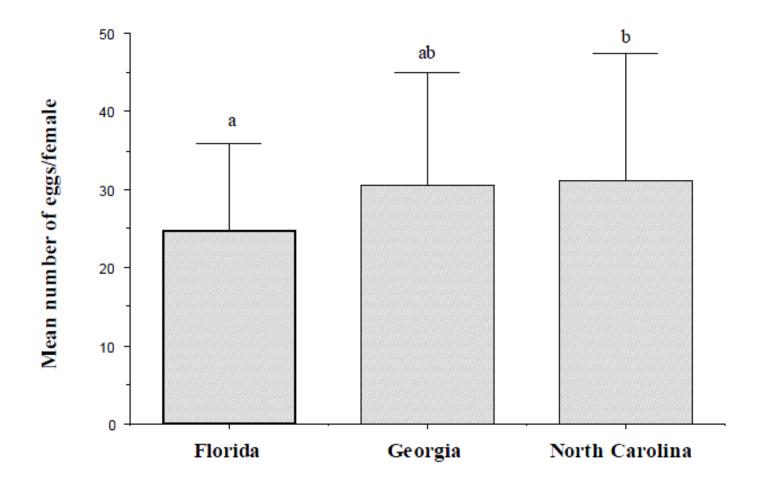


Figure 2. Clutch size for *W. smithii* collected from different locations

**Table 3:** Nei's Genetic Identity (above diagonal) and Genetic Distance (below diagonal) for three populations of *Wyeomyia smithii* (FL, GA, NC) and *Aedes aegypti* (AA) in 1998.

Population		Populations			
	FL	GA	NC	AA	
FL	-	0.88	0.65	0.04	
GA	0.13	-	0.72	0.02	
NC	0.44	0.32	-	0.19	
AA	3.11	3.75	1.67	-	



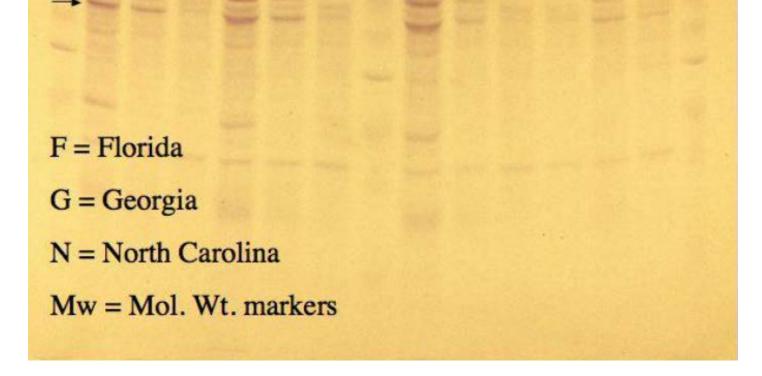
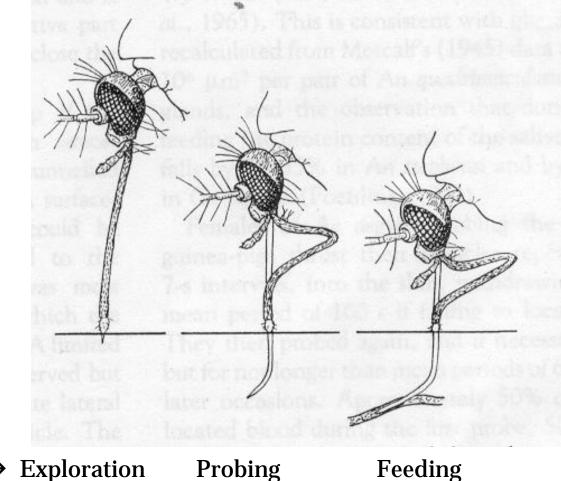


Figure 3. Differential hexamerin expression in adult female *W. smithii* 

# Overall results of *W. smithii* studies done in 1997-1998

- Georgia population is intermediate in all measures between Florida and North Carolina populations
- Georgia population is more similar genetically to Florida population
  - isozymes
  - hexamerin use
- Georgia population is more similar behaviorally and reproductively to NC population
  - almost complete lack of blood-feeding
  - similar reproductive output

#### Timed measures of blood-feeding stages: 2004-2008



Landing  $\rightarrow$  Exploration

Probing

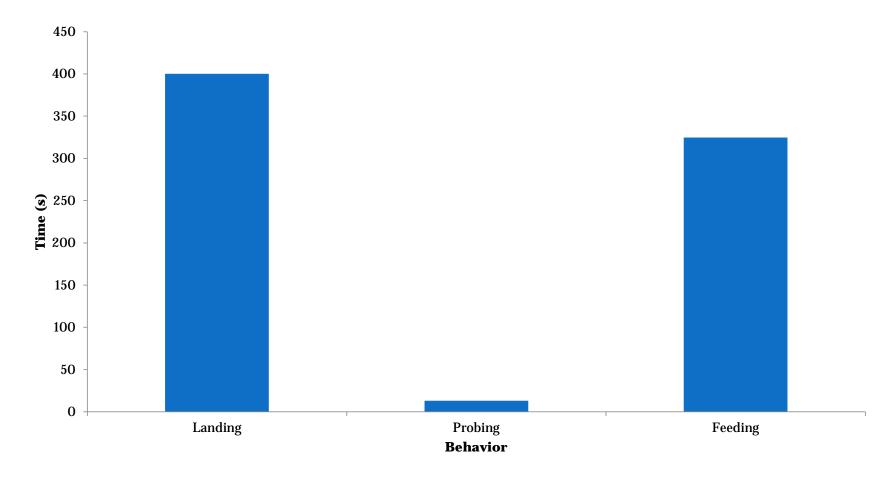


Figure 4. Mean duration of feeding behaviors of *W. smithii* in 2004

#### Table 4. Mean duration of blood feeding phases of W. smithii in 2004

Species	Host	Mean Duration of Phase (seconds)			
		Exploratory (Landing)	Probing	Feeding	
Wy smithii	Man	(400.1)	13.32	324.7	
Ae aegypti	Mouse	-	42	213	
	Guinea-pig	-	56	132	
	Man	3	68	220	
Ae africanus	Man	-	32	80	
Ae cinereus	Man	11	25	82	
Ae cantans	Man	8	28	150	
Cq richiardii	Man	16	92	162	
An plumbeus	Man	31	40	99	

Data other than for *W. smithii* from Clements, 1999.

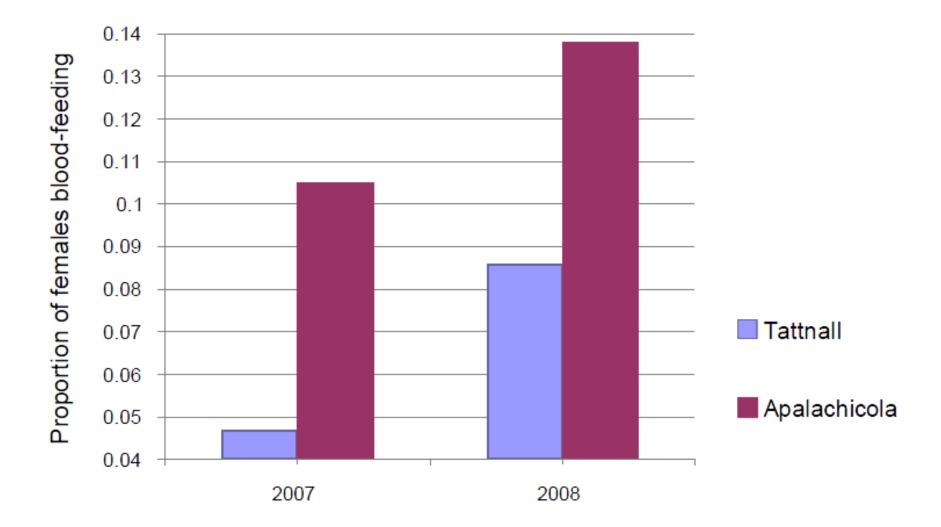


Figure 4. Results of cage blood feeding trials of *W. smithii* in 2007 and 2008

Table 5. Results of blood-feeding trials\* for post-autogenous adult female *Wyeomyia smithii* from 2004-2011

Population		Ave. % Feeding (± SD) by year			
	1998	2004	2007	2008	2011
Apalachicola	+**	ND***	8.0 (7.3)	13.8 (11.2)	26.0 (15.1)
Tattnall	±	2.3 (1.3)	3.8 (6.2)	8.6 (5.7)	20.8 (15.7)
Highlands	0	ND	0	0	0

\*6-12 15 minute feeding trials/population, 20-125 female mosquitoes/trial, all blood-fed specimens removed after each trial

\*\*In feeding trials with mosquitoes from Apalachicola, FL, mosquitoes were observed to blood-feed; in trials with Tattnall Co., GA mosquitoes, one female probed but did not feed \*\*\*not done

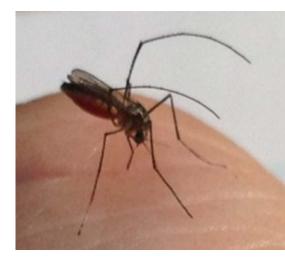
#### Overall results, 2004-2011:

- Georgia mosquitoes are increasing:
  - frequency of blood-feeding  $(1.1 \rightarrow 20.8\%)$
  - success at blood-feeding (albeit slowly):
    - 2004: partially engorged, <sup>1</sup>/<sub>4</sub> contain clear fluid
    - 2007: fully engorged, 1/10 contain clear fluid
    - 2008: fully engorged, all contain blood

Blood-feeding rate increasing in Florida as well

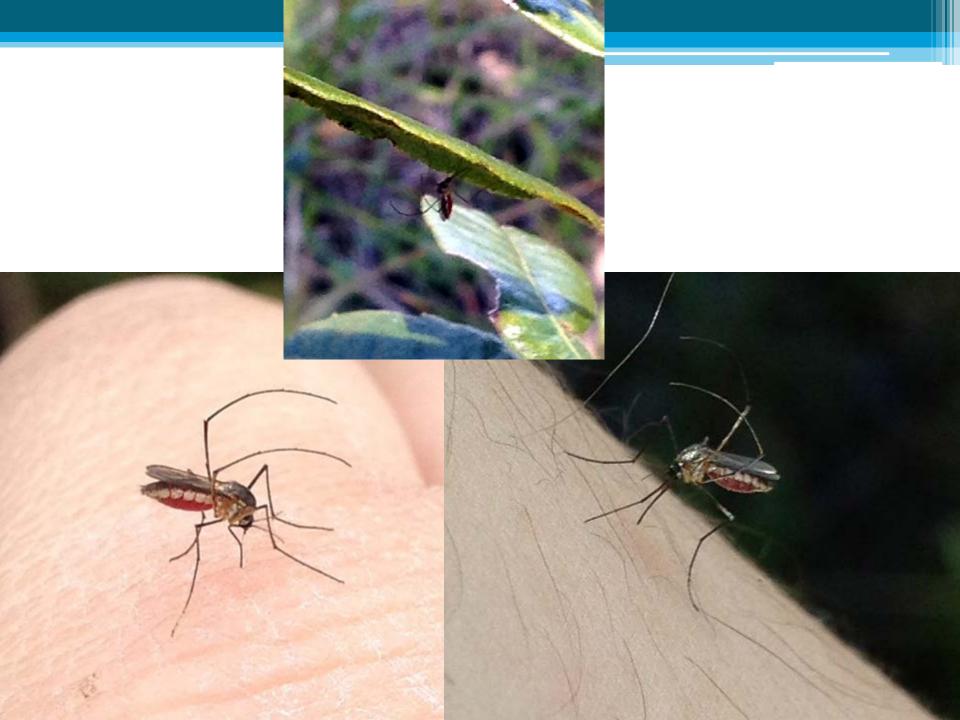
#### Current work: Bionomic Analysis

- Weekly surveys of larvae in pitchers
  Six pitchers from six rosettes
- Measured volume in each pitcher
- Counts of larvae at each instar
- Blood-feeding behavior recorded









#### **Bionomic Analysis Results**

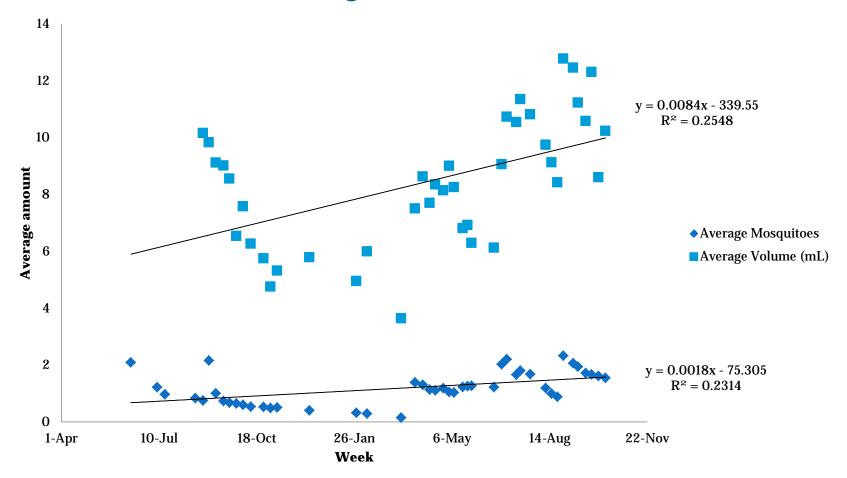


Figure 5. Regression of the average amounts of *W. smithii* larvae with the average volume in pitchers over time

#### **Bionomic Analysis Results**

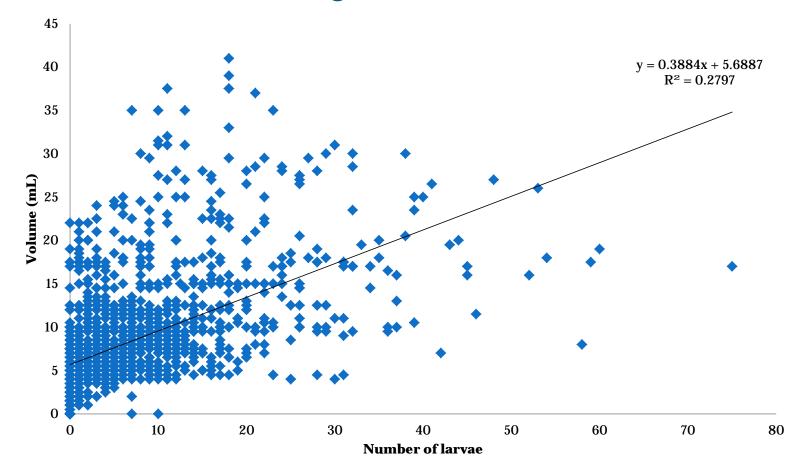


Figure 6. Correlation of the amount of *W. smithii* larvae with the volume of the pitcher

#### **Blood Feeding Behavior**

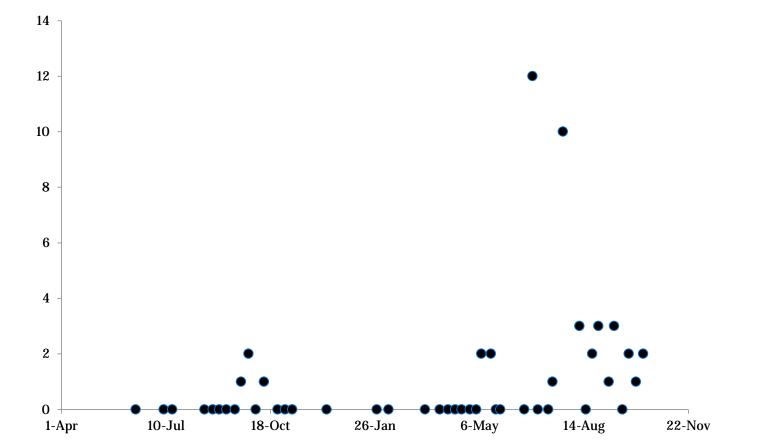
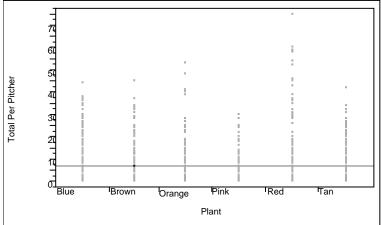


Figure 7. Number of times blood meals were taken by female W. smithii per week

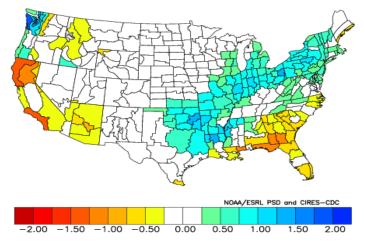
#### **Statistical Analyses**

- Spearman's Correlation
  - Spearman  $\rho$  0.7841, Prob> $|\rho| < 0.0001$
  - Strong correlation between volume of fluid in pitchers and number of mosquitoes present
- Kruskal-Wallis Test
  - H- 84.3877, DF- 5, Prob>H- <0.0001</p>
  - Distributions of larvae have different locations by rosettes.

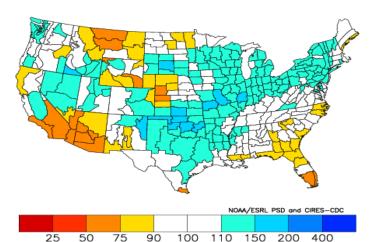


#### **Climatic Data**

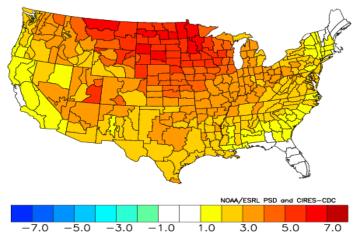
Composite Precipitation Anomalies (inches) Jan 1998 to 2007 Versus 1950–1995 Longterm Average

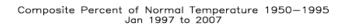


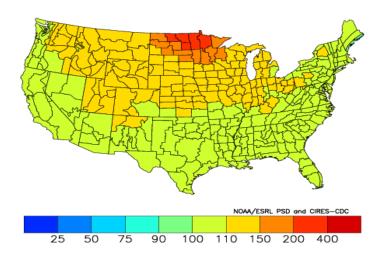
Composite Percent of Normal Precipitation 1950-1995 Jan 1997 to 2007



Composite Temperature Anomalies (F) Jan 1998 to 2007 Versus 1950–1995 Longterm Average







#### Population Genetics Analysis

- Assessed via isozyme analysis of isocitrate dehydrogenase (IDH) using polyacrylamide gel electrophoresis (PAGE)
- Evaluations between 6-12 from each of 9 plants
  - 4<sup>th</sup> instar larvae
  - Collected from field and reared at 25°C, 16L:8D



#### Population Genetics Results

Table 6. Observed heterozygotic frequencies and expectedheterozygotic frequencies of W. smithii

Subpopulation (S)	Samples (N)	Observed Heterozygotic Frequencies	Expected Heterozygotic Frequencies
S1	6	0	0
S2	12	0.08333	0.21875
S3	9	0.33333	0.27777
S4	9	0.44444	0.34567
S5	9	0.11111	0.10492
S6	12	0.16666	0.15277
S7	11	0.36666	0.29752
S8	9	0	0
S9	11	0.18888	0.16528

#### Population Genetics Results

Figure 8. Resolution of IDH -1 (monomorphic) and IDH -2 (polymorphic) alleles using PAGE

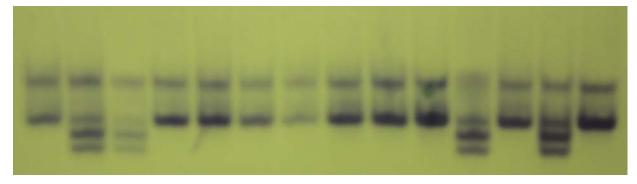


Table 7. F-statistics for comparison of population genetic structure for pitcher plant larvae within subpopulations (in pitchers within rosettes) and between rosettes

Site	F <sub>IS</sub>	F <sub>ST</sub>	F <sub>IT</sub>
Tattnall, GA	-0.057	0.049	-0.005

### Conclusions

# Low heterozygote frequencies for IDH F-statistics indicate high level of inbreeding

- Bionomics shows coincidental declines in larvae and pitcher volume
  - Reflective of low volume
  - Volume strongly correlated to total larvae
  - Distributions of larvae differ by plant
- Blood feeding behavior is increasing in the field
- Population of *W. smithii* is at risk
  - Long term drought and increasing temperatures
  - Reduction of S. purpurea

### Acknowledgements

- Dr. William Irby, advisor
- Phillip Bloodworth
- Kelly Dabney
- Casey Wesselman

